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Product Information

Citrate Buffer, pH 6.0, 10×
Antigen Retriever

Catalog Number **C9999**
Storage Temperature 2–8 °C

Synonym: Antigen Retrieval Solution

Product Description
Citrate buffer, pH 6.0, is used as a heat-induced antigen retriever on formalin-fixed paraffin-embedded (FFPE) tissue sections prior to application of antibodies.

In immunohistochemistry (IHC), most commonly used fixatives such as formalin mask tissue antigens (cellular, membrane, and nuclear) by their intrinsic crosslinking. This masking results in poor or no staining in IHC. The use of Citrate buffer, pH 6.0, or other antigen retrieval solutions on FFPE tissue sections improves accessibility of antibodies to tissue antigens.

Precautions and Disclaimer
This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Preparation Instructions
Dilute the Citrate buffer, pH 6.0, 10×, Antigen Retriever 10-fold with water to prepare a 1× Working Solution, e.g., dilute 10 mL of 10× concentrate with 90 mL of water.

Storage/Stability
Store the product at 2–8 °C. Do Not Freeze to prevent possible precipitation.
Note: Occasionally the buffer may contain a yellowish tinge. This is due to preservative and will not affect the function of the buffer.

Store the prepared 1× Citrate buffer, pH 6.0, Working Solution at 2–8 °C.

Procedure
The 1× Working Solution is intended for heat-induced antigen retrieval in IHC. Please refer to primary antibody protocol.

1. Deparafinize FFPE tissue section as outlined in the primary antibody protocol. Hydrate tissue with phosphate buffered saline.

2. Immerse the tissue slides in 1× Working Solution. Heat slides in solution for 10–20 minutes at 100 °C. Alternatively, heat slides in solution under pressure for 3–5 minutes. Remove and cool the slides at room temperature for 20 minutes.

3. The tissue sections are ready for further IHC staining.

Note: Tissue sections from brain or tissues with high concentration of lipids may become detached from slides or lose tissue morphology. For these tissues, it may be necessary to use positively charged, silanized, or poly-lysine coated slides.

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